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Efficacy of Fungicides Against *Alternaria Pluriseptata* and *Geotrichum Candidus* Incitant of IVY Gourd Fruit

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Abstract - IVY gourd suffers from pre and post-harvest diseases. This paper describes the sensitivity (MIC) of *Alternaria pluriseptata* and *Geotrichum candidus* against chlorothalonil and mancozeb fungicides. Sensitivity of 15 isolates of *A.pluriseptata* was tested against chlorothalonil. There was large variation in the sensitivity of these isolates. Some isolates were sensitive (600 µg/ml) while others were resistant (2500 µg/ml) i.e. ranged from 600 µg/ml to 2500 µg/ml while in case of mancozeb sensitivity ranged from 60 µg/ml to 1100 µg/ml . In case of *G.candidus* the sensitivity of 10 isolates were tested against chlorothalonil and mancozeb. In chlorothalonil some isolates were sensitive & resistant i.e. ranged from 650 µg/ml to 2000 µg/ml while in case of mancozeb sensitivity ranged from 20µg/ml to 300 µg/ml. *in vivo* results also showed positive.

Key words: *Coccinia indica*, *Alternaria pluriseptata* , *Geotrichum candidus*, fungicides

1. Introduction

IVY gourd (*Coccinia indicia* Wight & Arn.) of family Cucurbitaceae is distributed in tropical Asia, Africa, Pakistan, and India & Srilanka (Cook 1903, Sastri 1950). It is a climber & trailer (Nasir & Ali, 1973). The fruit is used as vegetable when green & eaten fresh when ripened. Every part of this plant is valuable in medicine. The IVY gourd, however suffers from pre & post harvest disease caused by *A.pluriseptat* (Karst &Har). Isolates & *Geotrichum candidus* (link: Leman) .The cultures were deposited at ASC College Naldurg and Agharkar Research Institute (ARI) Pune investigation was undertaken to evaluate the sensitivity of *A.pluriseptat* and *G.candidus* against Chlorothalonil & Mancozeb fungicides.

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2. Material and Methods

Sensitivity of isolates were tested against chlorothalonil & mancozeb determined by food poisoning test (Dekker and Gielink, 1979). Czapek Dox agar plates were prepared containing different concentration of fungicides mancozeb & chlorothalonil. Seven days fresh cultur disc (9&4mm) of the isolates were inoculated at the centre of plates in triplicate. The plates were incubated at 28±°c in the dark and radial growth was measured at different intervals. Plates without fungicides treated as control. *In vitro* studies, *A.pluriseptata* for chlorothalonil (100 to 2500 µg/ml) & for mancozeb (100 to 2000 µg/ml) concentrations were prepared. While in *G.candidus* for chlorothalonil (100 to 750 µg/ml) & mancozeb (10 to 300µg/ml) were prepared.

In vivo experiment was conducted on IVY gourd fruits by using the different concentrations *A.pluriseptata* chlorothalonil ranged from 100 to 3000 µg/ml & mancozeb ranged 10 to 1500 µg/ml. For *G.candidus* ranged from chlorothalonil 100 to 3000 µg/ml and mancozeb ranged 10 to 350 µg/ml were used.

Table: 1: Sensitivity of *Alteranaria pluriseptata* isolates from *Coccinia indica*

Against fungicide.

S N o.	Isolate	Place	Fungicides			
			Chlorothalonil (MIC) µg/ml (9mm disc)		Mancozeb (MIC) µg/ml (4mm disc)	
			<i>Invit ro</i>	<i>Inv ivo</i>	<i>Invit ro</i>	<i>Inv ivo</i>
1	Ap ₁	Naldurg	600 *	700*	120	130
2	Ap ₂	Murum	1500	1700	80	90
3	Ap ₃	Lohara	1600	1800	70	80
4	Ap ₄	Osmana bad	900	1200	90	100
5	Ap ₅	Omerga	1200	1400	400	400

6	Ap ₆	Nilanga	2000	2200	700	800
7	Ap ₇	Paranda	1400	1600	300	400
8	Ap ₈	Solapur	2000	2300	800	900
9	Ap ₉	Auranga bad	2000	2100	1100 +	1500 +
10	Ap ₁₀	Pune	1800	1900	500	600
11	Ap ₁₁	Beed	2500 +	3000 +	100	130
12	Ap ₁₂	Latur	1300	1500	60*	70*
13	Ap ₁₃	Mumbai	1700	1900	900	950
14	Ap ₁₄	Thane	1900	2000	300	400
15	Ap ₁₅	Jalna	1100	1400	1000	1200

* Sensitive + Resistant, Ap - *Alternaria pluriseptata* MIC - Minimum Inhibitory Concentration

Table 2: Sensitivity of *Geotrichum candidus* isolates from *Coccinia indica*

Against fungicide.

S N o.	Isolate	Place	Fungicides			
			Chlorothalonil (MIC) µg/ml (9mm disc)		Mancozeb(MIC) µg/ml (4mm disc)	
			In vitro	In vivo	In vitro	In vivo
1	Gc ₁	Mumbai	1300	1400	20*	50*
2	Gc ₂	Anala	1500	1600	50	60
3	Gc ₃	Solapur	650*	750*	90	100
4	Gc ₄	Latur	1600	1700	220	230
5	Gc ₅	Auranga bad	1500	1600	100	150
6	Gc ₆	Andur	1300	1400	140	150
7	Gc ₇	Omerga	1900	2000	270	280
8	Gc ₈	Naldurg	1800	1900	150	170
9	Gc ₉	Murum	2000 +	3000+	180	200
10	Gc ₁₀	Nilanga.	1800	1900	300+	350+

* Sensitive + Resistant, Gc- *Geotrichum candidus*, MIC - Minimum Inhibitory Concentration

3. Results and Discussion

This paper describes the sensitivity (MIC) of *A. pluriseptata* & *G.candidus* against chlorothalonil & mancozeb fungicides. Sensitivity of 15 isolates of *A. pluriseptata* was tested against chlorothalonil. There was large variation in the sensitivity of these isolates. Some isolates were sensitive (600µg/ml) i.e. Ap1 while others were resistant (2500µg/ml). i.e. Ap12, Ap6, Ap8, Ap9, Ap14. Its ranged from (600 to 2500 µg/ml). In case of mancozeb, sensitivity ranged from 60 µg/ml to 1100 µg/ml & sensitive isolates are Ap1, Ap2, Ap3, Ap4, Ap11, and Ap12 & tolerant isolates were Ap9, Ap13 and Ap15 (Table 1, Fig1&2).

Table 2 showed that the sensitivity of *G.candidus* 10 isolates in chlorothalonil, some isolates were sensitive isolates Gc3 (650 µg/ml) and highly tolerant isolates 2000 µg/ml i.e. Gc9 while in case of mancozeb, sensitivity ranged from 20 µg/ml to 300 µg/ml (Fig 3&4).

In vivo experiments, showed positive results. Sensitivity of *A. pluriseptata* against chlorothalonil ranged from 700 to 3000 µg/ml while in mancozeb its ranged from 50 to 400 µg/ml. In case of *G. candidus* against chlorothalonil ranged from 750 to 3000 µg/ml while in mancozeb it was 50 to 350 µg/ml.

Variation in the sensitivity of different pathogens in relation to many fungicides have been reported (Jones and Ehret, 1976, Dekker & Gielink, 1979, Gangawane & Saller, 1981; Gangawane and Shaikh, 1988; Hollomon, 1981, Kamble, 1991, Bhale, 2002). Annamalai and Lalithakumari (1996) suggested that it is essential to establish the base line sensitivity for the fungicide against sensitive strain, Brain (1980) considers that heterogeneous population of nuclei consisting of resistant and sensitive nuclei in the isolates might be responsible for variation in the MIC of fungicides. Recently Bhale and Gogle reported the development of carbendazim resistance in *Alternaria spinaciae* incitant of spinach (*spinacia oleracea* L.). There was variation in MIC of Ridomil Gold among the five isolates of *Phytophthora palmivora* var. *piperina* on the agar plates (Patil and Kamble, 2011).

References

1. Annamalai, P. and Lalithakumari, D. 1996. Decreased Sensitivity of *Drechslera oryzae*. Field isolates to edifenphos, Ind. Phytopath. 43(4): 553-558.
2. Bhale, U.N. 2002. Studies on Management of some important Diseases of Spinach in Maharashtra. Ph.D. thesis: Dr. Babasaheb Ambedkar Marathwada University, Aurangabad.
3. Bhale, U.N. & D.P. Gogle (2008). Effect of passage on the Development of carbendazim Resistance in *Alternaria spinaciae* incitant of leaf spot of spinach (*Spinacea oleracea* L.). GEOBIOS. 35: 37-40.
4. Brain, G.C.A. 1980. Resistance in peronosporale to acylalanine type fungicides. Ph.D. thesis, Univ. Guelph, Ontario, Canada, pp 10.

5. Cooke, C.I.E.T. Flora of Presidency of Bombay, Vol 1. Published under the Authority of Secretary of state for Council, 1903.
6. Dekker J. and Gielink A.J. 1979. Acquired resistance to pimarinin in *Cladosporium cucumerinum* and *Fusarium oxysporum* sp. narcissi associated with decreased virulence. Neith. J. Plant Pathol. 85:67-73.
7. Gangawane, L.V. and R.S. Saler, 1981. Resistance to fungicides in *Aspergillus flavus*. Neth. J. Pl.Path. 87-254.
8. Gangawane, L.V. and Shaikh, S.A. 1988. Development of resistance to aluminium ethyl phosphate in *pythium aphanizomatense*. Indian phytopathology. 41(41) 638-641.
9. Hollomon, D.W. 1981. Mode of action of hydroxy pyrimidine fungicides. Fungicides Resistance in protection (Dekker, J. and S.G. Geogopoulos Eds.) CAPD wegingen, Netherlands.
10. Jones, A.L. and Ehret, G.R. 1976. Tolerance to fungicides in *Venturia* and *Monilinia* of tree fruits. Proc. Ann. Phytopath. Soc. 3:84-90.
11. Nasir, E. and Ail, S.I. Flora of West Pakistan, Cucurbitaceae, No. 154, Botany Department, University of Karachi, 1973.
12. Patil, K.B. and Kamble, S.S. (2011). Efficacy of Ridomil Gold Against *Phytophthora palmivora* Var. *piperina* causing Quick wilt of Black Pepper. Bioinfolet, 8 (1) : 105-106
13. Sastri, B.N. (1950). The wealth of India, A Dictionary of Raw materials and industrial products. Publication and information of Directorate CSIR, New Delhi Vol 2 & 8, pp. 257 & 285-293.

**In vitro**

2500 µg/ml(R)

600 µg/ml(S)

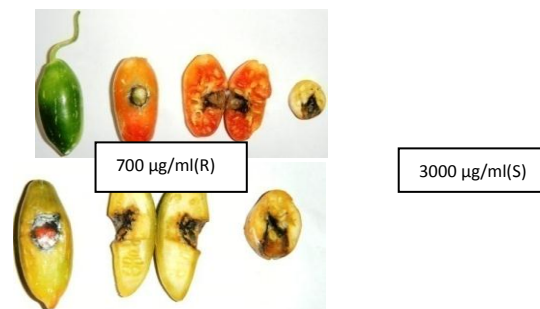
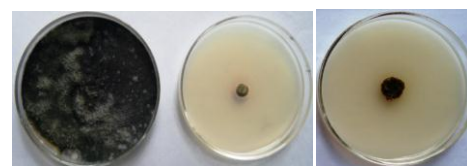
**Fruits**

Figure 1 - Sensitivity of *Alteranaria pluriseptata* isolates from *Coccinia indica* against fungicides chlorothalonil.



CONTROL

1100 µg/ml(R)

60 µg/ml(S)

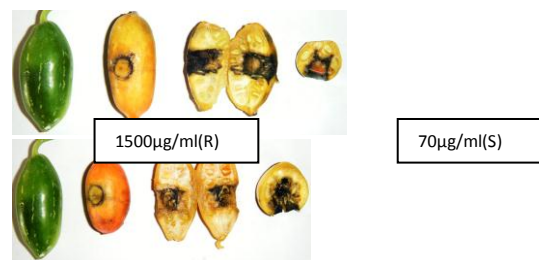
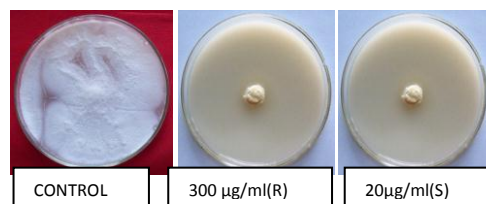
**Fruits**

Figure 2 - Sensitivity of *Alteranaria pluriseptata* isolates from *Coccinia indica* against fungicides mancozeb.



CONTROL

300 µg/ml(R)

20 µg/ml(S)

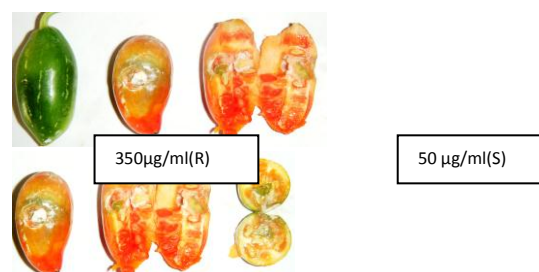
In vitro**Fruits**

Figure 3 - Sensitivity of *Geotrichum candidus* isolates from *Coccinia indica* against fungicides chlorothalonil.

350 µg/ml(R)

50 µg/ml(S)

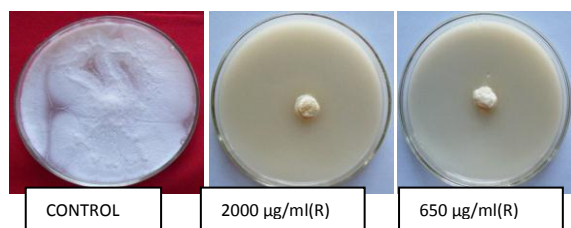
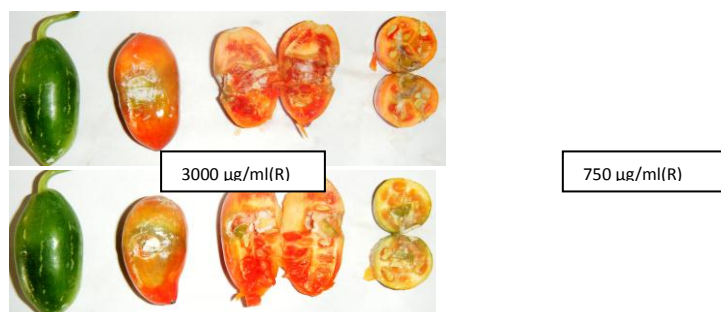
**In vitro****Fruits**

Figure 4 - Sensitivity of *Geotrichum candidus* isolates from *Coccinia indica* against fungicides mancozeb

Tolerance of Polluted Water on Seedling Growth of Some Cereal Crops

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Abstract - Attempts were made to study the influence of sugar mill industrial polluted water on seed germination and seedling growth of cereal crops from Tuljabhavani Sahakari Sakhar Karkhana Naldurg. The higher concentration of distillery effluent were found to inhibit the germination of cereal crops. Maize (*Zea mays*) and Rice (*Oryza sativa*) were more susceptible and Jowar (*Sorghum vulgare*) was less susceptible to polluted water as compared to other crops. Polluted water showed low pH (2.02) and high temp., OD, TS, TDS & TSS than tap water. In the phytotoxicity was more in *Sorghum vulgare* than other crops.

Key words: Effluent, pollution, seed germination, cereal crop seeds

1. Introduction

The various kinds of pollution caused by rapid industrialization. The problem of water pollution due to industrial effluents waste water. The most important effluent discharging industries are distilleries, sugar mills, Pulps, paper mills etc. Polluted water not only affects animal life but also the vegetations. An investigation was conducted to utilize and observe the influence of Tuljabhavain Sahakari Sakhar Karkhana Naldurg polluted water on seed germination and seedling growth of some important cereal crops.

2. Materials and Methods

Five different types of cereal crops seeds viz. *Penisetium typhoides* (Bajara), *Sorghum vulgare* (Jowar), *Zea mays* (Maize), *Oryza sativa* (Rice) and *Triticum aestivum* (Wheat) were selected for the present experiment. Thirty seeds of each were soaked in polluted water and another set in tap water as control for 24 hr in separate Petri dishes. Using filter paper method (Agrawal, 1995), the seeds were folded in germinating paper. Each paper contains 10 seeds in triplicate. The folded paper along with seeds were tied with thread and kept in 500 ml beaker. 200 ml polluted water was poured in it, while another beaker was kept as control by using tapwater. After every two days interval, water was added as per requirements depending upon rate of absorption. After seven days the germinated seeds were removed. Shoot & Root length, fresh and dry weight of the seedlings were measured & analyzed statistically (Mungikar, 1997). The standard method of (APHA, 1992), (Saxena, 1990), and (Trivedy,

1998) were followed for the analysis of effluent.

The phytotoxicity (%) was calculated using the formula Chou et al. (1978).

Percentage of phytotoxicity

$$= \frac{\text{Length of Control} - \text{Length of Test (Polluted)}}{\text{Length of Control}} \times 100$$

The Effluent tolerance index (ETI) was calculated using the formula determined by Turner & Marshal (1972).

Effluent tolerance index

$$= \frac{\text{Mean length of the longest root \& shoot in the Effluent}}{\text{Mean length of the longest root \& shoot in the control}}$$

3. Result and Discussion

Table -1. Analysis of physico-chemical parameters of tap water and polluted water used for seedling growth.

Parameter	Tap water	Polluted water
Colour	Transparent	Faint yellow colour.
Odour	No specific smell.	Foul smell.
Temp.(°c)	29	32
pH	7.2	2.02
Optical Density (OD)	0.01	0.67
Total Solids (TS) mg/l.	320	550
Total dissolved Solids. (TDS) mg/l	300	500
Total Suspended Solids. (TSS) mg/l.		150
		250

Table 2. Effect of polluted effluent on seed germination of some cereal crops.

Sr.No.	Characterstics		Bajra	Jowar	Maize	Rice	Wheat
1.	Seed Germination (%)	Control	96.00	86.00	23.00	96.00	91.00
		Polluted	04.00	10.00	92.00	73.00	04.33
2.	Root length (cm)	Control	01.2±0.4	01.0± 0.2	02.99±0.3	82.23±0.1	04.99±0.3
		Polluted	00.00	0.30 ± 0.2	04.11 ±0.3	03.32±0.4	00.10 ± 0.1
3.	Shoot length (cm)	Control	04.11 ±0.2	02.22 ± 0.2	03.33±0.2	07.22±0.1	05.11±0.2
		Polluted	00.00	00.00	3.22±0.4	4.11±0.2	1.00±0.1
4.	Fresh weight of root (mg)	Control	00.70	00.70	01.32	00.21	00.10
		Polluted	00.00	00.10	00.30	00.09	00.10
5.	Fresh weight of Shoot (mg)	Control	00.20	00.60	01.11	00.20	00.770
		Polluted	00.00	00.300	00.255	00.210	00.20
6.	Dry weight of root (mg)	Control	00.17	00.34	00.00	00.10	00.200
		Polluted	00.00	00.10	00.40	00.22	00.002
7.	Dry weight of Shoot (gm)	Control	00.10	00.10	00.00	00.50	00.23
		Polluted	00.00	00.20	00.50	00.10	00.20
8.	Phytotoxicity (%)	Root	16.80	71.90	00.00	13.00	17.33
		Shoot	00.00	33.30	00.00	31.50	12.33
9.	Effluent tolerance index (%)	Root	00.00	00.20	00.60	28.60	12.33
		Shoot	00.00	00.45	00.60	04.80	09.14

The physical properties of industrial water used for seedling are given in table 1. The presence of total suspended solids to the extent of 250 mg/l was responsible for faint yellow coloration of waste water. The pH of Polluted water 2.02, total dissolved solids (500 mg /l) total Solids (550 mg / l) & optical density (0.67) was stunted seedling growth. Moreover root also adversely affected the fresh and dry weight of all seedlings except *Z. mayes* and *O. sativa* in polluted water.

The shoot length, root length, fresh and dry weight of different seedling treated with polluted water and tap water are give in Table 2. The observations showed that *Z. mayes* and *O. sativa* were most susceptible to polluted water as compared to others. Its root length and shoot

length were decreased that form 4.11 to 2.99, 3.32 to 2.23 & 3.22 to 3.33, 4.11 to 7.22 cm respectively due to polluted water, while its fresh weight increased to 0.300 gm and 0.255 gm which were 1.32 gm & 1.11 gm respectively in control. Seed in polluted has maximum germination i.e. 92.00%, 73.00% in *Z.mayes* and *O.sativa*. Seed in polluted has minimum 04.00% in *S.vulgare* and *P.typhoides*. Phytotoxicity was increased in shoot of *O.sativa* (31.50 %) and effluent tolerance index was also adversely affected in root and shoot.

The results showed that the industrial polluted water adversely affected seedling growth of cereal crops due to suspended solid and low pH of polluted water. Therefore, it is concluded that the industrial polluted

water from "Tuljabhabani Sahakari Sakhar karkhana Naldurg" is suitable for seedling growth of *Z.mayes* and *O.sativa*.

Several workers like Arokia Sami et. al. (1981) , Sahai et. al. (1983), Gupta et al. (2003), Swaminathan & Vidheeswaran (1997), Nanda et al,(1991) & Shreshtha & Niroula (2003) have observed the influence of different kinds of industrial effluents on seed germination and seedling growth. The higher concentration of distillery effluent were found to inhibit the germination and growth of paddy (*Oryza sativa L.*) (Suresha et.al,2006). Population studies of sugar mill effluent showed that most of the physico - chemical parameters like colour, odour total solids, COD, BOD, alkalinity and fluoride were found to be exceed the ISI prescribed permissible values while pH , phosphate and sulphate were found within permissible limit (Senthil Kumar et.al; 2001). Recently, Bhale and Gogle (2008) were reported the *Hibiscus esculentus* was more susceptible and *vigna radiata* was more resistant to polluted water as compared to other vegetable seeds.

References

- [1] **Agrawal, R. L.** (1995). *Seed Technology 2nd ed.*, Oxford and IBH publishing Co. New Delhi.
- [2] **APHA.** (1992). *Standard Methods for the Examination of water and Waste water. 19th ed.* Washinton D. C., U.S. A.
- [3] **Arokia Sami, D.I., Meenakshi, R.M. and Gnanarethinan, J.L.** (1981).
- [4] Effect of distillery effluent on water potential of rice seedlings, *Indian J. exp. BioL*, 19, 96-98.
- [5] **Bhale, U.N. and Gogle,D.P.**(2008). Impact of industrial polluted water on seedling growth of vegetable crops.*Geobios*,35:41-42.
- [6] **Chou, C. S. Chang, C. & Kaw, C. I.** (1978). Impact of water pollution of crop growth in Taiwan. *Bot. Bull, Bot, Sinica*, 19,107-124.
- [7] **Gupta, R., Gupta, Rudra and Shukla** (2003), D.N. Effect of Sewage water on seed germination and seedling growth of *Vigna sinensis* and *Abelmoschus esculentus*. *Proc. of the Nat. Acad. of Sci. India, Section B*, 73-82.
- [8] **Mungikar, A.M.** (1997). An introduction to Biometry. *Saraswati Printing press, Aurangabad*.
- [9] **Nanda, B., Tripathy, S.K. and Padhi , S.** (1991), Effect of algalization on seed germination of vegetable crops. *J. Microb. Biotech.*, 7, 622-623.
- [10] **Sahai, R., Jabeen, S. and Sazena, P. K.** (1983). Effect of distillery waste on seed germination, seedling growth and pigment contents of rice. *Indian J. Ecol.* 10, 7-10.
- [11] **Saxena, M.M.** (1990). The Analysis of Water, Air and Soil BSI and ISI Standards. *Meerut Publishere*.
- [12] **Senthil Kumar, R.D. Narayanaswamy R. and Ramakrishnan, K.** (2001). Pollution Studies on sugar mill effluent, physico - chemical Characteristics toxic metals. *Pollution Res.* 20 (1): 93 - 97.
- [13] **Suresha, G. & E.T. Puttaih.** (2006). Effect of distillery effluent on seed germination growth of paddy (*Oryza sativa*) *Geobios.* 33; 129-132.
- [14] **Shrestha, M.K. and Niroula, B.** (2003) Germination behaviour of pea seeds on municipality sewage and some industrial effluents of Biratnagar. *Our Nature*, 1, 33-36.
- [15] **Swaminathan, K. and Vaidheeswaran, S.** (1991) Effect of dyeing factory effluents on seed germination and seedling development of groundnut (*Arachis hypogaea*). *Environ. Biol.*, 12, 353-358.
- [16] **Trivedy, R. K. Goel, P.K. and C.L. Trishal,** (1998). *Practical methods in Ecological And Environmental Sciences.* Media publication, Karad, India.
- [17] **Turner, R. C. & Marshal, C.** (1972). Accumulation of Zink by subcellular fraction of some root Agrotics teneyes in relation to zink tolerance. *New Phyton.* 71, 671-676.